

by treating 12.0 g (2.95 mmol) of 7 for 2, 2, and 25 min with 1:2 (v/v) TFA/CH₂Cl₂ containing 1:49 (v/v) ethanedithiol, followed by five 2-min washes with CH₂Cl₂. Neutralization was carried out with 1:9 (v/v) triethylamine/CH₂Cl₂ for periods of 2, 2, and 4 min, followed by five 2-min washes with CH₂Cl₂ and one DMF wash. Coupling of the fragment was performed by treating the neutralized resin with a solution of the activated N-terminal fragment prepared as follows: a stirred solution of 6.9 g (5.27 mmol) of 5 and 606 mg (5.27 mmol) of *N*-hydroxysuccinimide (recrystallized) in 15 mL of degassed DMF was cooled in an ice-water bath for 5 min, and 5.27 mL of a 1 M solution of DCC in CH₂Cl₂ was added in one portion. The ice bath was removed, and the thick mixture was stirred for 4 h. After adding this solution to the resin, 159 mg (1.3 mmol) of recrystallized (ethyl acetate) 4-(dimethylamino)pyridine (DMAP) was added, and the mixture was rocked gently for 22 h. Then 1.0 mL of DCC solution was added and mixing continued for 26 h at which time a solution of 159 mg (1.3 mmol) of DMAP in 3.0 mL of degassed DMF was added and mixing continued for an additional 64 h. The resin was washed twice with DMF for 2 min each, followed by six 2-min washes with CH₂Cl₂. After removing the Boc protection using the TFA solution described above for 3- and 25-min periods followed by six 2-min washes with CH₂Cl₂, the peptide-resin was dried to constant weight at reduced pressure to afford 19.5 g (91%) of 2: aaa avg = 0.138 mmol/g, Arg (Orn) 5.047 (5), Asx 2.285 (2), Glx 1.157 (1), Gly 5.010 (5), Ala 1.120 (1), Ile 1.685 (2), Leu 0.962 (1), Tyr 1.056 (1), Phe 1.678 (2).

Arg-Arg-Ser-Ser-Cys(Acm)-Phe-Gly-Gly-Arg-Ile-Asp-Arg-Ile-Gly-Ala-Gln-Ser-Gly-Leu-Gly-Cys(Acm)-Asn-Ser-Phe-Arg-Tyr-OH (8). A Kel-F reaction vessel was charged with a slurry of 2.0 g (0.28 mmol) of 2 and 1.0 g (6.7 mmol) of *L*-methionine in 4.0 mL of *m*-cresol (Aldrich), which was stirred slowly for 1 h. The reaction vessel was attached to a Kel-F manifold and cooled in a dry ice/acetone cold bath for approximately 10 min, and approximately 45 mL of anhydrous HF (Matheson) was condensed into the stirred mixture. The cold bath was replaced with an ice-water bath, the mixture was stirred in the cold for 75 min, and the HF was removed in the cold under reduced pressure (water aspirator for 70 min followed by vacuum pump for 1 h). After triturating the residue with 50 mL of ether for 15 min in the cold, the mixture was filtered and washed with two 30-mL portions of ether and dried briefly under reduced pressure. The peptide was leached from the resin by stirring and filtering with 15 mL of 1:1 (v/v) acetic acid/H₂O. Purification was achieved by applying the combined filtrates onto a Sephadex G-25F column (5 × 100 cm) and eluting with 2.0 M acetic acid. Product purity was checked by assaying the fractions (22 mL) by TLC (system IX) and HPLC (system II, column A). Fractions of appropriate purity were combined and evaporated to dryness at reduced pressure. Material from four identical runs was lyophilized from 150 mL of H₂O to afford 1.765 g (41%) of 8: aaa

avg = 0.259 mmol/g, Arg (Orn) 4.918 (5), Asx 2.038 (2), Ser [3.476] (4), Glx [1.164] (1), Gly [4.369] (5), Ala 1.052 (1), Ile 1.968 (2), Leu 1.032 (1), Tyr 1.017 (1), Phe 1.976 (2); TLC (system X) *R*_f 0.3; HPLC (system II, column A) 86%, *t*_R 50 min.

Arg-Arg-Ser-Ser-Cys-Phe-Gly-Gly-Arg-Ile-Asp-Arg-Ile-Gly-Ala-Gln-Ser-Gly-Leu-Gly-Cys-Asn-Ser-Phe-Arg-Tyr-OH, Disulfide Form (1). A solution of 1.39 g (5.5 mmol) of I₂ in 620 mL of 80% HOAc was added rapidly to a briskly stirred solution of 1.21 g (0.3 mmol) of 8 in 37 mL of 50% HOAc. After 2 h the reaction mixture was cooled in an ice-water bath for 5 min and treated with 4.9 g of zinc dust until decolorization of the I₂ was complete. The mixture was filtered, and the filtrate was concentrated at reduced pressure to a volume of about 25 mL, diluted with an equal volume of H₂O, and charged onto a 5 × 100 cm column of Sephadex G-50F in 50% HOAc. The column was eluted with 50% HOAc, and the fractions were pooled on the basis of HPLC analysis (system II, column A), combined, concentrated at reduced pressure, and lyophilized from H₂O to give 0.92 g of crude 1. A solution of 1.76 g of this material in 50 mL of 0.05 M NH₄OAc (pH adjusted to 5.0 with acetic acid) was applied to a 300-mL (4.0 cm diameter) column of carboxymethylcellulose (CMC) (Whatman) equilibrated with 0.3 M NH₄OAc (prepared by diluting 76.5 mL of concentrated NH₄OH (29% NH₃) and 68.6 mL of glacial acetic acid to 4.0 L with degassed H₂O and acidifying to pH 5.0 with additional acetic acid). The column was eluted with 4 L of 0.3 M NH₄OAc (pH 5.0, prepared as above) and 22-mL fractions were collected over a period of 24 h. Those fractions showing greater than 97% purity by HPLC (system II, column A) were combined, applied directly to a Sephadex G-25F column (5 × 100 cm), and eluted with 2.0 M acetic acid to remove NH₄OAc. The fractions containing product were combined, evaporated to dryness under reduced pressure, and lyophilized from 125 mL of H₂O to give 890 mg (40.6%) of 1 as a colorless solid: aaa avg = 0.286 mmol/g, Arg 5.04 (5), Asx 2.05 (2), Ser 4.06 (4), Glx 1.00 (1), Gly 4.90 (5), Ala 1.00 (1), Ile 1.93 (2), Leu 1.02 (1), Tyr 1.00 (1), Phe 2.00 (2); TLC (system X) *R*_f 0.33; HPLC (system II, column A) 97.3%, *t*_R 49 min; GC, acetic acid 6.9%; H₂O, Karl Fischer titration 8.1%; specific rotation [α]_D²⁶ +47.3°; cumulative sequence preview ≤3% at Ser³⁰ cycle. Combustion analysis (based on tetraacetate minus water). Calcd: C, 49.17; H, 6.66; N, 19.42; S, 2.07. Found: C, 49.11; H, 6.80; N, 19.32; S, 2.42.

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New Approaches to the Synthesis of *trans*-Alkene Isosteres of Dipeptides

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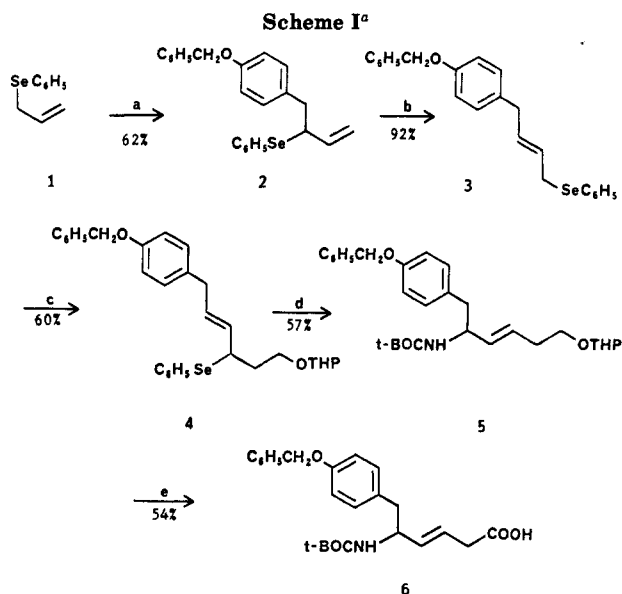
Two new syntheses of protected dipeptide analogues bearing a *trans* carbon-carbon double bond in place of the amide linkage are reported. One route is a linear synthesis employing the rearrangement of an allylic selenide to a protected allylic amine. The second route is convergent and uses the Julia olefin synthesis in a key step. The latter route is fully stereocontrolled and has been used to prepare protected *trans*-alkene isosteres of the dipeptides TyrAla, PhePhe, LeuPhe, and LeuLeu.

Introduction

Studies of structure-function relationships in the field of biologically active peptides have largely focused on the effects of changes in side chain residues. This is experimentally advantageous, since peptide synthesis is now routine, but does not permit the study of questions regarding the role played by the amide backbone. Changing

the amide backbone is in general more synthetically challenging, but, despite this, numerous amide replacements have been pursued.¹ It has been suggested that the

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^a (a) (i) LDA, THF, -78 °C, (ii) LDA, C₆H₅CH₂O-*p*-C₆H₄CH₂Br; (b) PTSA-H₂O, C₆H₆, reflux; (c) (i) LDAz, THF, -78 °C, (ii) THP-OCH₂CH₂Br; (d) *t*-C₄H₉OCONH₂, Et₃N, NCS, CH₃OH, 0 °C; (e) (i) 0.2 M HCl (aqueous), THF; (ii) Jones oxidation.

replacement of the amide linkage with a *trans* carbon-carbon double bond provides a substance which closely mimics the three-dimensional shape of the parent amide but that is inert to enzymatic hydrolysis. It has further been predicted that peptide analogues bearing the so-called *trans*-alkene isostere will retain biological activity if the amide linkage so replaced is not involved in the secondary or tertiary structure of the peptide or in the mechanism whereby it elicits its biological response.² One reviewer has called the *trans*-alkene isostere "an ideal replacement".¹

When this project was initiated, there existed two procedures for the preparation of dipeptide analogues containing the *trans*-alkene isostere.^{2,3} It has been shown that these dipeptide mimics could be incorporated into larger peptides. Both routes successfully afforded *trans*-alkene isostere-containing mimics of enkephalin;^{2,3} one of the routes also provided a substance P analogue.³ Subsequently these methods were used to prepare an angiotension-converting enzyme inhibitor⁴ and several renin inhibitors^{5,6} containing this replacement. Most of these compounds showed promising biological activity.

Both of these synthetic routes suffered from lack of stereocontrol with respect to either or both the alkene linkage and the side chain-bearing stereocenters. We envisioned developing a sequence which would surmount these difficulties. Two new synthetic routes to protected *trans*-alkene isosteres of dipeptides have been developed

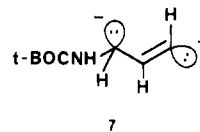
and are reported herein.⁷ One of these routes is the first which is fully stereocontrolled.

Results and Discussion

An Organoselenium Route. *trans*-Alkene isosteres contain an embedded allylic amine. The sigmatropic rearrangement of oxidatively activated allylic selenides which yields allylic amines⁸ was developed for this application and was used in the preparation of *trans*-alkene isosteres; such a synthesis of the protected TyrGly isostere 6 is illustrated in Scheme I.

The anion of allyl phenyl selenide (1)⁸ was alkylated⁹ with *p*-(benzyloxy)benzyl bromide, and the resulting allylic selenide 2 was treated in refluxing benzene with a trace of *p*-toluenesulfonic acid to afford 3, the product of allylic rearrangement. Allylic selenide 3 was again alkylated to yield 4, which was in turn subjected to NCS-promoted rearrangement⁸ in the presence of *tert*-butyl carbamate, to yield protected allylic amine 5. Although the THP ether could be removed and the resulting alcohol oxidized in a single step with Jones reagent,¹⁰ a more reproducible procedure involved hydrolysis of the THP ether¹¹ followed by Jones oxidation.

It is noteworthy that the acid-catalyzed rearrangement of allylic selenides, as in 2 → 3, has been previously noted¹² but has not to our knowledge been used as a synthetic reaction. This alkylation–rearrangement–alkylation–rearrangement sequence makes allyl phenyl selenide the synthetic equivalent of the dianion 7.



The organoselenium approach to *trans*-alkene isosteres (Scheme I) is stereocontrolled with respect to the geometry of the alkene⁸ but suffers from several drawbacks. Although in principle extendable to the synthesis of peptide mimics with residues other than Gly at the C-terminus, this has proven nontrivial, since the requisite alkylation of 3 with a more substituted halide (for example to directly prepare a TyrAla analogue) could not be achieved. An indirect route involving esterification of a Gly-C-terminal acid (such as 6), followed by alkylation at the α -carbon¹³ and finally saponification, is thus required. A second limitation is that this route does not, in the present form, address the question of stereocontrol of the two chiral centers. A final drawback is the linearity of the sequence, which renders inconvenient the preparation of gram quantities of final isostere by this method. A new route was sought.

An Organosulfur Route. It appeared that the two major flaws with the organoselenium route, linearity and lack of stereocontrol, might be eliminated by a connective scheme such as the Julia olefin synthesis.¹⁴ The simplest

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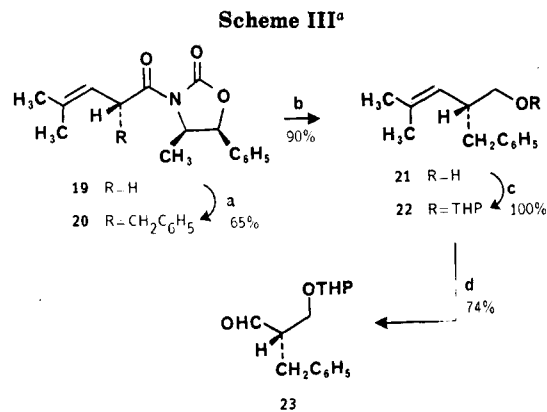
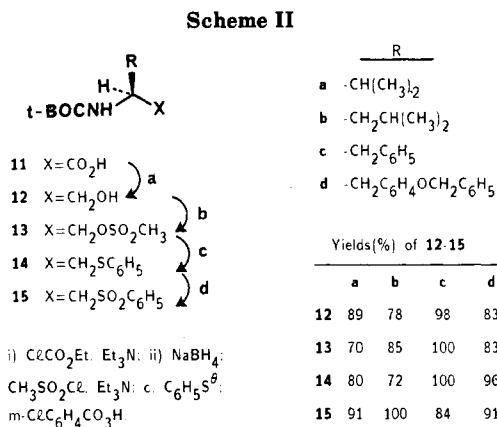
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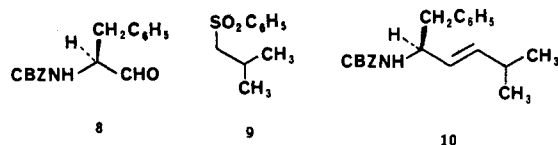
(13) Cox^{3a} has achieved alkylations on closely related compounds.



^a (a) (i) LDA, THF, -78 °C, (ii) C₆H₅CH₂Br; (b) LiAlH₄, THF, -78 → 0 °C; (c) DHP, PyrH⁺OTS⁻, CH₂Cl₂, 25 °C; (d) (i) O₃, -78 °C, (ii) (CH₃)₂S.

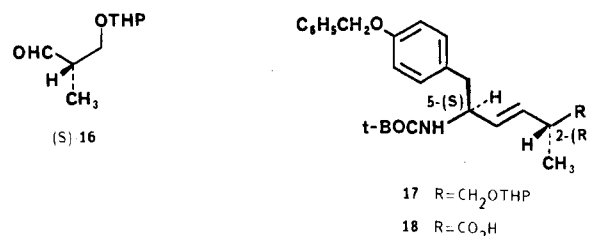
option was coupling of a protected α -amino aldehyde, available by several methods,¹⁵ with a sulfone anion appropriately functionalized to serve as the eventual C-terminus.

Attempts to couple CBZ-phenylalanyl 8 and the anion of isobutyl phenyl sulfone (9) followed by sodium amalgam reduction¹⁶ were discouraging, providing, at most, traces of olefinic product 10, despite extensive variation of reaction conditions. Fortunately, developments along another line allowed us to abandon this approach.



A connective approach in which the N-terminus enters as a sulfone anion and the C-terminus as an aldehyde was more successful. N-protected α -amino acids are readily converted to the requisite N-protected β -amino sulfones by a four-step sequence in which all steps proceed in high yield (Scheme II^{17,18}) and chromatography is not required, since the final products can be purified by recrystallization. The *t*-BOC protected β -amino sulfones 15 are stable indefinitely at 25 °C.

The coupling reaction was studied by using the sulfone 15d derived from tyrosine and aldehyde (S)-16, derived from commercially available methyl 3-hydroxy-2(S)-methylpropionate by protection as the THP ether (dihydropyran, pyridinium tosylate, CH₂Cl₂), followed by reduction with DIBAL. When the dianion of 15d, prepared in THF at -78 °C with methyllithium, was treated with 2 equiv of aldehyde (S)-16 and the crude mixture of β -hydroxy sulfones was reduced with sodium phosphate-buffered methanolic sodium amalgam, the yield of alkene 17 was poor (~20%). The problem appeared not to be the reduction step but rather the coupling. Trapping of the β -oxido sulfone was considered as a solution. Mixtures of the dianion of 15d and (S)-16 were thus treated with



Better results were obtained in the presence of diisobutylaluminum methoxide. In practice, a solution of 2 equiv of DIBAL in THF at 0 °C was treated with 2 equiv of methanol, cooled to -78 °C, followed by addition of 2 equiv of aldehyde (S)-16. The resulting mixture was added to 1 equiv of the dianion of 15d, prepared at -78 °C by the addition of 2 equiv of methyllithium to 1 equiv of 15d. It was found that the aldehyde/aluminum complex is unstable even at -78 °C and is best used within a few minutes of its preparation. Reduction with sodium amalgam followed by chromatography on silica gel afforded the alkene (2R,5S)-17 in 63% overall yield. Direct oxidation¹⁰ of (2R,5S)-17 afforded the protected *trans*-alkene isostere of TyrAla, (2R,5S)-18.

A simple technical modification of the above procedure is possible when the C-terminal aldehyde is to be prepared by DIBAL reduction of the corresponding ester. In this case, the DIBAL reduction reaction mixture (in ether) can be directly used in the coupling to the sulfone anion. This method avoids the need to isolate the aldehyde component and, in the case of (2R,5S)-17, provided a superior yield (73%).

To exclude the possibility of racemization at either of the stereocenters of 18, the *trans*-alkene isostere synthesis was repeated with 15d and (R)-16, affording (2S,5S)-18. The methyl doublets in the ¹H NMR spectra of the two diastereomers were clearly resolved at 500 MHz (2R,5S)-18 (δ 1.25; (2S,5S)-18, δ 1.22 as a mixture) and conclusively demonstrated that appreciable racemization had not occurred during any step in the synthesis of 18.

The use of protected β -amino sulfones for the synthesis of protected allylic amines is new. It is interesting and not necessarily expected that only the *trans* isomer of the product was detected.¹⁴ The presence of a negative charge on the nitrogen is apparently important for the success of this reaction, preventing β -elimination of the nitrogen to yield a vinyl sulfone. Consistent with this was the failure of the *t*-BOC-protected β -aminosulfone derived from proline to undergo coupling under the above conditions,

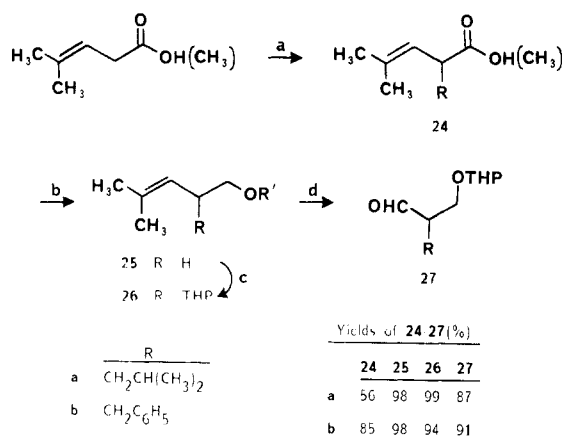
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Scheme IV^a

^a (a) (i) LDA, THF, 0 °C, (ii) RX, -78 → 25 °C; (b) LiAlH₄, Et₂O; (c) DHP, PyrH⁺OTs⁻, CH₂Cl₂, 25 °C; (d) (i) O₃, CH₂Cl₂, (ii) (C-H)₂S.

affording instead what was identified as a vinyl sulfone. The β-elimination could be suppressed for the *t*-BOC-protected sulfone derived from proline by carrying out the metalation and coupling at -110 °C, but yields of alkene were quite low (~20%).

The luxury of commercial availability of a direct precursor to the optically active C-terminal aldehyde is unique to alanine. A general route to optically active aldehydes for incorporation at the C-terminus of *trans*-alkene isosteres was found in Evans alkylation¹⁹ of oxazolidone **19** followed by multistep functional group manipulation as shown in Scheme III. The indicated absolute configuration of **21** was verified by chemical correlation with the known (*R*)-2-benzyl-3-(benzyloxy)propanol.²⁰ C-terminal phenylalanine equivalent **23** was prepared in this manner. Racemic aldehydes such as **27a,b**, C-terminal leucine and phenylalanine equivalents, respectively, were available by an analogous route from the illustrated β,γ-unsaturated carboxylic acid (or ester) (Scheme IV).

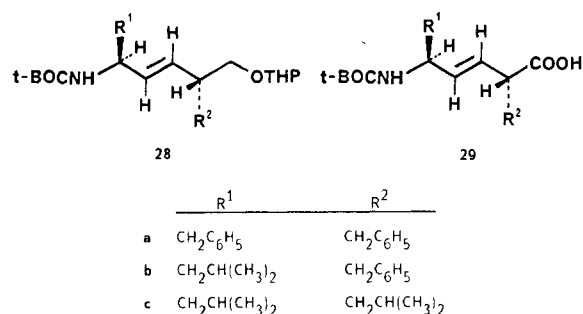
Table I illustrates the structures and yields of the *trans*-alkene isosteres prepared by various combinations of the protected β-amino sulfones and protected β-hydroxy aldehydes.

Conclusion

Application of the Julia olefin synthesis of *trans*-alkene isosteres has afforded the first fully stereocontrolled approach to these interesting peptide mimics. The route has provided sufficient quantities of the PhePhe and LeuPhe isosteres to allow testing of some new renin inhibitors.⁶ It is possible that the route described herein could be applied to the synthesis of more functionalized *trans*-alkene isosteres bearing side chains such as Ser, Asp, Lys, etc., but this will certainly require judicious choice of protective groups.

Experimental Section²¹

3-(Phenylseleno)-4-[(4-phenylmethoxy)phenyl]butene, Allylic Selenide 2. *n*-Butyllithium (12.7 mL, 33 mmol, 2.6 M in hexanes) was added to a 0 °C solution of diisopropylamine (3.54 g, 35 mmol) in 80 mL of THF, and the mixture was stirred 0.5

Table I. Synthesis of *trans*-Alkene Isosteres of Dipeptides

		yield, %	
protected β-amino sulfone	protected β-hydroxy aldehyde	THP ether	acid
15d	(<i>S</i>)- 16	17, 63 (73 ^a)	18, 64
15c	(<i>S</i>)- 23	28a , 37	29a , 68
15b	(<i>S</i>)- 23	28b , 65	29b , 63
15b	(±)- 27a	28c , 62 ^b	29c , 65, 60 ^c

^a Method 2, see Experimental Section. ^b Mixture of diastereoisomers at C-2. ^c Diastereoisomers **28c** separated and independently oxidized.

h and cooled to -78 °C. Allyl phenyl selenide (**1**) (6.34 g, 32.2 mmol) was added over 5 min to produce a deep yellow solution, which was stirred 0.5 h at -78 °C and then treated with 7.5 g (27 mmol) of *p*-(benzyloxy)benzyl bromide, added as a solid, and maintained at -78 °C for 2 h. The mixture was quenched with 4 mL of water and diluted with 50 mL of ether. Drying (MgSO₄) and concentration in vacuo afforded the product as an orange oil, which was chromatographed on silica gel (4% ethyl acetate/hexanes) to yield 6.6 g (62%) of selenide **2** as a yellow oil: ¹H NMR (500 MHz) δ 2.94 (1 H, dd, *J* = 6 and 8 Hz, ArCH₂), 3.01 (1 H, dd, *J* = 6 and 7 Hz, ArCH₂), 3.92 (1 H, m, SeCH), 4.70 (1 H, d, *J* = 19 Hz, C=CH), 4.81 (1 H, d, *J* = 9 Hz, C=CH), 5.05 (2 H, s, OCH₂), 5.83 (1 H, m, CH=C), 6.9 (5 H, m, C₆H₅), 7.1–7.5 (9 H, m, Ar); LRMS, *m/e* 394, 392, 237, 91; IR (NaCl) 3027, 1595, 1490, 1000, 875, 715 cm⁻¹.

1-(Phenylseleno)-4-[(4-phenylmethoxy)phenyl]-*trans*-2-butene, Allylic Selenide 3. A solution of 1.75 g (4.5 mmol) of selenide **2** and 75 mg (0.44 mmol) of *p*-toluenesulfonic acid in 200 mL of benzene was stirred for 3 h at reflux. The mixture was cooled to 25 °C and quenched with 30 mL of saturated aqueous NaHCO₃. The organic layer was dried (MgSO₄), concentrated in vacuo, and chromatographed on silica gel (5% ethyl acetate/hexanes) to afford selenide **3** as a yellow oil, 1.61 g (92%): ¹H NMR (500 MHz) δ 3.23 (2 H, d, *J* = 6 Hz, CH₂Ar), 3.51 (2 H, d, *J* = 6 Hz, CH₂Se), 5.05 (2 H, s, CH₂O), 5.51 (1 H, dt, *J* = 18 and 8 Hz, HC=C), 5.62 (1 H, dt, *J* = 18 and 8 Hz, C=CH), 6.9–7.5 (14 H, m, Ar); LRMS, *m/e* 394, 392, 237, 91; IR (NaCl) 3025, 1595, 1490, 1000, 945 cm⁻¹.

4-(Phenylseleno)-1-[4-(phenylmethoxy)phenyl]-6-[(2-tetrahydropyranyl)oxy]-*trans*-2-hexene, Allylic Selenide 4. Allylic selenide **3** (12.4 g, 31.5 mmol) was metalated with LDA (34 mmol) and alkylated with the THP ether of 2-bromoethanol (7.5 g, 36.0 mmol) as described above for the preparation of selenide **2**. Allylic selenide **4** was isolated by chromatography on silica gel (8% ethyl acetate/hexanes), 9.84 g (60%), as a yellow oil: ¹H NMR (500 MHz) δ 1.5–1.7 (6 H, m, THP), 2.0 (2 H, m, OCH₂CH₂), 3.21 (2 H, d, *J* = 6 Hz, ArCH₂), 3.45 (2 H, m, OCH₂), 3.83 (2 H, m, CH₂O), 3.92 (1 H, m, CHSe), 4.55 (1 H, m, OCHO), 5.05 (2 H, s, ArCH₂O), 5.33 (1 H, m, C=CH), 5.50 (1 H, m, CH=C), 6.9–7.5 (14 H, m, Ar); LRMS, *m/e* 522, 365, 281, 263, 197, 91, 65; IR (CHCl₃) 3020, 2310, 1590, 1480, 880 cm⁻¹.

2-[(*tert*-Butoxycarbonyl)amino]-1-[4-(phenylmethoxy)phenyl]-6-[(2-tetrahydropyranyl)oxy]-*trans*-3-hexene, Protected Allylic Amine 5. Selenide **4** (1.3 g, 2.5 mmol), *tert*-butyl carbamate (0.87 g, 7.5 mmol), and triethylamine (2.0 g, 19.7 mmol) in 2.5 mL of CH₃OH was cooled to 0 °C and treated with 1.0 g (7.5 mmol) of *N*-chlorosuccinimide over 10 min. The resulting suspension was stirred 0.5 h at 0 °C, diluted with ethyl acetate, and washed sequentially with 10 mL of water and 10 mL

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(21) General procedures are described elsewhere.⁸

of saturated aqueous NaCl. Drying (MgSO_4) and concentration in vacuo gave a black oil, which was chromatographed on silica gel (20% ethyl acetate/hexanes) to yield protected allylic amine **5**, 0.69 g (57%), as a colorless solid: $^1\text{H NMR}$ (500 MHz) δ 1.3 (9 H, s, $t\text{-C}_4\text{H}_9$), 1.5–1.7 (6 H, m, CH_2O), 2.7 (2 H, m, ArCH_2), 3.3 (2 H, m, OCH_2), 3.5–3.7 (2 H, m, CH_2O), 4.3 (2 H, m, CHNH), 4.6 (1 H, m, OCHO), 5.05 (2 H, s, ArCH_2O), 5.6 (2 H, m, $\text{CH}=\text{CH}$), 6.9–7.5 (9 H, m, Ar); LRMS, m/e 398, 144, 91, 85, 68, 65, 57; IR (CHCl_3) 3300, 1711, 1590, 1480 cm^{-1} .

Protected Racemic TyrGly Isostere 6. THP ether **5** (0.42 g, 0.87 mmol) in 4.0 mL of 0.2 M aqueous HCl and 15 mL of THF was heated for 2.5 h at 55 °C, cooled to 25 °C, diluted with 20 mL of ethyl acetate, and washed sequentially with 10 mL of saturated aqueous NaHCO_3 and saturated aqueous NaCl. The organic layer was dried (MgSO_4) and concentrated in vacuo to afford 0.33 g (95%) of the alcohol as a colorless oil, which solidified on standing: $^1\text{H NMR}$ (80 MHz) δ 1.4 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.2 (2 H, m, allylic CH_2), 2.7 (2 H, m, ArCH_2), 3.5 (2 H, m, OCH_2), 4.3 (2 H, m, CHNH), 5.05 (2 H, s, ArCH_2), 5.4 (2 H, m, $\text{HC}=\text{CH}$), 6.8–7.4 (9 H, m, Ar); LRMS, m/e 281, 280, 237, 200, 197, 144, 100, 91, 83, 65, 57.

The alcohol (50 mg, 0.125 mmol) was oxidized as described below for the preparation of the protected LeuPhe isostere **29b** to afford TyrGly isostere **6**, 29 mg (57%), as a colorless solid: $^1\text{H NMR}$ (500 MHz) δ 1.4 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.75 (2 H, d, ArCH_2), 3.09 (2 H, d, allylic CH_2), 4.40 (2 H, m, NHCH), 5.05 (2 H, s, ArCH_2), 5.60 (2 H, m, $\text{CH}=\text{CH}$), 6.89 and 7.10 (4 H, m, Ar), 7.35 (5 H, m, Ar).

2(S)-[(*tert*-Butoxycarbonyl)amino]-3-[4-(phenylmethoxy)phenyl]propanol (12d). A solution of 15.0 g (40.4 mmol) of *t*-BOC-*O*-benzyl-L-tyrosine (**11d**) and 5.56 mL (50 mmol) of triethylamine in 50 mL of THF was cooled to –5 °C. Ethyl chloroformate (3.86 mL, 40.4 mmol) was added dropwise during 0.25 h. The resulting slurry was stirred at –5 °C for 0.5 h and filtered. The residue was washed with 20 mL of THF, and the combined filtrates were added dropwise to a slurry of 3.82 g (100 mmol) of NaBH_4 in 50 mL of water at 0 °C. The reaction mixture was stirred for 4 h at 0 °C and extracted with ether and CH_2Cl_2 . Drying (MgSO_4) and concentration in vacuo (50 °C) afforded a white solid, which could be used without further purification or recrystallized (20% ethyl acetate–hexanes) to yield 13.13 g (91%) of the protected amino alcohol **12d** in two crops as a white solid: mp 108 °C; $^1\text{H NMR}$ (500 MHz) δ 1.42 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.25 (1 H, s, OH), 2.77 (2 H, d, $J = 7$ Hz, ArCH_2C), 3.54 (1 H, dd, $J = 5$ and 11 Hz, CH_2OH), 3.66 (1 H, dd, $J = 3$ and 11 Hz, CH_2OH), 3.82 (1 H, m, CHCH_2OH), 4.69 (1 H, s, NH), 5.04 (2 H, s, CH_2OAr), 6.7–7.4 (9 H, m, Ar); IR (CHCl_3) 3620, 3440, 3080, 3040, 2980, 2940, 2860, 1702, 1682, 1610, 1510, 1500, 1392, 1369, 1240, 1163, 1205, 865 cm^{-1} ; LRMS, m/e 357 (M^+), 301, 284, 270, 240, 226, 198, 197, 160, 130, 127, 124, 115, 107, 104, 101, 91 (100), 86, 85, 84, 77, 71, 69, 60, 51; $[\alpha]^{20}_{\text{D}} -17.0^\circ$ (c 0.06 g/mL, CHCl_3).

2(S)-[(*tert*-Butoxycarbonyl)amino]-1-[(methylsulfonyl)oxy]-3-[4-(phenylmethoxy)phenyl]propane, Mesylate 13d. A solution of 2.50 g (7.0 mmol) of the protected amino alcohol **12d** and 2.1 g (21 mmol) of triethylamine in 50 mL of CH_2Cl_2 was cooled to 0 °C and treated dropwise with 1.96 g (17.1 mmol) of methanesulfonyl chloride followed by stirring for an additional 0.33 h at 0 °C. Addition of water and extraction with CH_2Cl_2 followed by drying (MgSO_4) and concentration in vacuo afforded crude mesylate **13d** as a yellow solid. Recrystallization (25% ethyl acetate–hexanes) yielded 3.05 g (100%) of mesylate **13d** as a white solid: mp 103–105 °C; $^1\text{H NMR}$ (500 MHz) δ 1.42 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.79 (2 H, m, ArCH_2C), 3.00 (3 H, s, CH_3), 4.11 (2 H, m, CH_2OSO_2), 4.22 (1 H, m, CHN), 4.69 (1 H, s, NH), 5.04 (2 H, s, ArCH_2O), 6.9–7.4 (9 H, m, Ar); IR (CHCl_3) 3440, 3040, 2980, 2940, 2865, 1760, 1710, 1612, 1585, 1505, 1452, 1370, 1240, 1178, 1028, 978 cm^{-1} ; LRMS, m/e 318 ($\text{M}^+ + t\text{-C}_4\text{H}_9\text{OCONH}_2$), 283, 198, 197, 138, 137, 107, 105, 96, 92, 91 (100), 89, 86, 85, 79, 78, 77, 65, 59, 57, 56; $[\alpha]^{20}_{\text{D}} -10.4^\circ$ (c 0.02 g/mL, CHCl_3).

2(S)-[(*tert*-Butoxycarbonyl)amino]-1-(phenylthio)-3-[4-(phenylmethoxy)phenyl]propane, Sulfide 14d. A solution of 1.30 g (25 mmol) of NaOCH₃ and 2.64 mL (25.9 mmol) of benzenethiol in 3.75 mL of CH_3OH and 18.75 mL of THF was stirred at 25 °C for 0.25 h. Mesylate **13d** (3.42 g, 7.85 mmol), was added as a solid, and the mixture was heated to 50 °C for 2.5 h. The solution was cooled to 25 °C and diluted with 10% aqueous

NaOH. Extraction with CH_2Cl_2 and drying (MgSO_4), followed by concentration in vacuo (40 °C), afforded a yellow solid, which was recrystallized (10% ethyl acetate–hexanes) to yield 2.9 g (84%) of sulfide **14d** as a white solid: mp 94 °C; $^1\text{H NMR}$ (500 MHz) δ 1.42 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.82 (2 H, m, ArCH_2), 3.06 (2 H, m, SCH_2), 4.02 (1 H, m, CHCH_2S), 4.64 (1 H, s, NH), 6.9–7.5 (14 H, m, Ar); IR (CHCl_3) 3440, 3080, 3060, 3020, 2980, 2930, 2860, 1705, 1609, 1552, 1510, 1496, 1452, 1438, 1368, 1297, 1242, 1168, 1043, 1025, 863 cm^{-1} ; LRMS, m/e 449 (M^+), 376, 359, 283, 270, 252, 227, 226, 223, 197, 196, 153, 152, 135, 99, 97, 92, 91 (100), 86, 85, 84, 81, 77, 69, 65, 59, 57; $[\alpha]^{20}_{\text{D}} +9.9^\circ$ (c 0.1 g/mL, CHCl_3).

2(S)-[(*tert*-Butoxycarbonyl)amino]-3-[4-(phenylmethoxy)phenyl]-1-(phenylsulfonyl)propane, Sulfone 15d. A solution of 2.90 g (6.45 mmol) of sulfide **14d** in 50 mL of CH_2Cl_2 was cooled to 0 °C and treated with 3.56 g (20.6 mmol) in *m*-chloroperoxybenzoic acid. The resulting slurry was stirred 1 h at 25 °C and then partitioned between CH_2Cl_2 and saturated aqueous NaHSO_3 /10% aqueous NaOH. The aqueous layer was extracted several times with CH_2Cl_2 , and the combined organic extracts were dried (MgSO_4). Concentration in vacuo afforded **15d** as a white solid, which was usually used in the next step without further purification. Recrystallization from methanol yielded 2.83 g (91%) of the sulfone **15d** as a white solid: mp 207–209 °C; $^1\text{H NMR}$ (500 MHz) δ 1.35 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.85–3.05 (2 H, m, ArCH_2C), 3.2–3.45 (2 H, m, CH_2SO_2), 4.08 (1 H, m, CH-N), 4.88 (1 H, br s, NH), 5.03 (2 H, s, ArCH_2O), 6.9–7.9 (14 H, m, Ar); IR (CHCl_3) 3440, 3390, 3040, 3010, 2980, 2840, 1710, 1690, 1610, 1510, 1450, 1370, 1320, 1310, 1280, 1245, 1150, 1088, 1050, 1025 cm^{-1} ; LRMS, m/e 481 (M^+), 446, 425, 408, 364, 309, 308, 273, 248, 235, 229, 228, 198, 197, 184, 173, 172, 128, 110, 107, 91 (100), 82, 78, 77, 72, 57; $[\alpha]^{20}_{\text{D}} -29.1^\circ$ (c 0.02 g/mL, THF); Anal. C, H, N.

Sulfones 15a-c. Sulfones **15a-c** were prepared from the appropriate *t*-BOC-protected L-amino acid by the four-step sequence described above in detail for **11** → **15d**.

2(S)-[(*tert*-Butoxycarbonyl)amino]-3-methyl-1-(phenylsulfonyl)butane, sulfone 15a: mp 103–105 °C (33% ethyl acetate–hexanes); $^1\text{H NMR}$ (80 MHz) δ 0.86 (3 H, d, $J = 7$ Hz, CH_3), 0.89 (3 H, d, $J = 7$ Hz, CH_3), 1.41 (9 H, s, $t\text{-C}_4\text{H}_9$), 1.75–2.21 (1 H, m, CH), 3.25 (2 H, m, CH_2SO_2), 3.61–3.95 (1 H, m, CHNH), 4.66 (1 H, br s, NH), 7.43–7.64 and 7.81–7.96 (5 H, m, Ar); IR (KBr) 3494, 2972, 1688, 1516, 1453, 1372, 1313, 1300, 1249, 1172, 1148, 1088, 880, 758, 698 cm^{-1} ; $[\alpha]^{20}_{\text{D}} +11.02$ (c 0.023 g/mL, CHCl_3).

2(S)-[(*tert*-Butoxycarbonyl)amino]-4-methyl-1-(phenylsulfonyl)pentane, sulfone 15b: mp 95–98 °C (33% ethyl acetate–hexanes); $^1\text{H NMR}$ (80 MHz) δ 0.88 (6 H, d, $J = 7$ Hz, $2 \times \text{CH}_3$), 1.24–1.79 (3 H, m, CHCH_2), 1.39 (9 H, s, $t\text{-C}_4\text{H}_9$), 3.32 (2 H, m, CH_2SO_2), 3.79–4.17 (1 H, m, CHNH), 4.71 (1 H, br s, NH), 7.44–7.65 and 7.79–7.96 (5 H, m, Ar); IR (KBr) 3484, 2962, 1692, 1614, 1449, 1365, 1288, 1172, 1149, 1079, 785, 749 cm^{-1} ; $[\alpha]^{20}_{\text{D}} -9.87^\circ$ (c 0.024 g/mL, CHCl_3).

2(S)-[(*tert*-Butoxycarbonyl)amino]-3-phenyl-1-(phenylsulfonyl)propane, sulfone 15c: mp 215–216 °C (50% CHCl_3 -ethyl acetate); $^1\text{H NMR}$ (80 MHz) δ 1.37 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.99 (2 H, d, CH_2), 3.25 (2 H, m, CH_2SO_2), 3.88–4.30 (1 H, m, CHNH), 4.80 (1 H, br s, NH), 7.04–7.29 (5 H, m, Ar), 7.45–7.64 and 7.77–7.91 (10 H, m, Ar); IR (KBr) 3388, 2980, 1692, 1520, 1448, 1368, 1287, 1173, 1149, 1138, 1088, 752, 707 cm^{-1} ; $[\alpha]^{20}_{\text{D}} -32.7^\circ$ (c 0.0052 g/mL, Me_2SO).

Methyl 3-[(2-Tetrahydropyranyl)oxy]-2(S)-methylpropionate. A solution of 5.33 g (45.1 mmol) of (*S*)-methyl 3-hydroxy-2-methylpropionate, 4.93 g (58.7 mmol) of 3,4-dihydropyran, and 0.22 g of pyridinium *p*-toluenesulfonate in 75 mL of CH_2Cl_2 was stirred at 25 °C for 3 h. The mixture was extracted sequentially with saturated aqueous NaHCO_3 and saturated aqueous NaCl and dried (MgSO_4). Concentration in vacuo (50 °C) afforded the THP ether as a colorless liquid (9.12 g, 100%): $^1\text{H NMR}$ (500 MHz) δ 1.18 and 1.19 (3 H, $2 \times$ d, $J = 7$ Hz, ratio 1:1, CH_3), 1.47–1.90 (6 H, m, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.79 (1 H, m, CHCOO), 3.4–3.95 (4 H, m, $2 \times \text{OCH}_2$), 3.695 and 3.69 (3 H, $2 \times$ s, ratio 1:1, OCH_3), 4.59 (1 H, m, OCHO); IR (NaCl) 201 ($\text{M}^+ - 1$), 184, 171, 147, 129, 119, 115, 102, 101, 88, 86, 85 (100), 84, 73, 69, 67, 56, 55 cm^{-1} ; $[\alpha]^{20}_{\text{D}} +17.1^\circ$ (c 0.11 g/mL, CHCl_3).

2(S)-Methyl-3-[(2-tetrahydropyranyl)oxy]propionaldehyde, (S)-16. A solution of 9.0 g (44 mmol) of the ester from

the preceding reaction in 150 mL of CH_2Cl_2 was cooled to -78°C , treated dropwise with 50.0 mL (47.5 mmol) of DIBAL (0.95 M in hexane), stirred at -78°C for 10 min, and quenched with saturated aqueous NH_4Cl . Filtration, drying (MgSO_4), and concentration of the organic layer in vacuo (50°C) afforded 9.16 g of crude aldehyde (S)-16 as a liquid. This material was normally used in the next step without further purification. For full characterization of the compound, 4.95 g of the above sample were chromatographed on silica gel (30% ethyl acetate-hexanes) to yield 3.57 g (87%) of the aldehyde (S)-16 as a colorless liquid: $^1\text{H NMR}$ (500 MHz) δ 1.15 and 1.16 (3 H, 2 \times d, $J = 7$ Hz, ratio 1:1, CH_3), 1.45–1.85 (6 H, m, $\text{CH}_2\text{CH}_2\text{CH}_2$), 2.63 (1 H, m, CHCHO), 3.48–3.62 and 3.78–4.00 (2 H, m, CH_2O), 4.58 (1 H, m, OCHO), 9.83 (1 H, m, CHO); IR (NaCl) 2940, 2870, 2720, 1720, 1452, 1441, 1386, 1352, 1261, 1202, 1120, 1078, 1062, 1040, 972, 903, 872, 818, 754 cm^{-1} ; LRMS, m/e 172 (M^+), 171, 115, 102, 101, 88, 86, 85 (100), 84, 71, 67, 59, 57, 56, 55; $[\alpha]_D^{20} +28.0^\circ$ (c 0.053 g/mL, CHCl_3).

5(S)-[(tert-Butoxycarbonyl)amino]-2(R)-methyl-6-[4-(phenylmethoxy)phenyl]-1-[(2-tetrahydropyranyl)oxy]-trans-3-hexene, THP Ether 17. Method 1. A solution of sulfone 15d (100 mg, 0.208 mmol) in 3 mL of THF at -78°C was treated over 5 min with 0.25 mL (0.458 mmol) of CH_3Li (1.8 M in ether) and stirred 0.33 h. In a separate flask, 72 mg (0.416 mmol) of aldehyde (S)-16 in 0.5 mL of THF at -78°C was treated with 0.42 mL of diisobutylaluminum methoxide (1 M in THF; prepared by the addition of 1 equiv of methanol to a 1.0 M solution of DIBAL in THF). The solution of the aluminum complex was transferred via cannula into the solution of the sulfone anion, and the whole was stirred 0.5 h at -78°C . The reaction was quenched at -78°C with saturated aqueous NH_4Cl and was extracted with ether, dried (MgSO_4), and concentrated in vacuo. The crude mixture of β -hydroxy sulfone diastereoisomers was dissolved in 3 mL of methanol, cooled to 0°C , and treated sequentially with Na_2HPO_4 (0.17 g, 3 mmol) and 1.7 g (10 mmol) of 5% Na (Hg). The mixture was stirred 4 h at 0°C , diluted with water, extracted with CH_2Cl_2 , and dried (MgSO_4). Concentration in vacuo followed by chromatography on silica gel (10% ethyl acetate-hexanes) afforded 64 mg (63%) of 1 as a colorless oil. Spectroscopic data were the same as described in the following procedure.

5(S)-(tert-Butoxycarbonylamino)-2(R)-methyl-6-[4-(phenylmethoxy)phenyl]-1-[(2-tetrahydropyranyl)oxy]-trans-3-hexene, THP Ether 17. Method 2. Methyl 3-[(2-tetrahydropyranyl)oxy]-2(S)-methylpropionate, (4.025 g, 20 mmol) in 60 mL of ether was cooled to -78°C , treated dropwise with 21.9 mL (21.9 mmol) of DIBAL (1 M in ether), and stirred for 0.5 h to yield solution A. In a separate flask, a solution of sulfone 15d (4.0 g, 8.3 mmol) in 100 mL of THF at -78°C was treated over 5 min with 11.0 mL (17.6 mmol) of CH_3Li (1.6 M in ether) and stirred 0.5 h. The latter sulfone anion solution was treated with solution A added via cannula over 0.25 h, and the whole was stirred 0.25 h at -78°C . The reaction was quenched at -78°C with saturated aqueous NH_4Cl , and the product was extracted with ether, dried (MgSO_4), and concentrated in vacuo. The crude mixture of β -hydroxy sulfone diastereoisomers was dissolved in 150 mL of CH_3OH and cooled to 0°C . Disodium hydrogen phosphate (13 g, 78 mmol) and 50 g (100 mmol) of 5% Na (Hg) were added sequentially. The mixture was stirred 4 h at 0°C , diluted with water, and extracted with CH_2Cl_2 and the organic layer dried (MgSO_4) and concentrated in vacuo. Chromatography on silica gel (40% ethyl acetate-hexanes) afforded 2.98 g (73%) of the THF-protected alcohol 17 as a colorless oil, which solidified upon standing at 25°C : $^1\text{H NMR}$ (500 MHz) δ 0.99 (3 H, m, CH_3), 1.2–1.9 (6 H, m, THP ether), 1.42 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.41 (1 H, m, $\text{CHC}=\text{C}$), 2.77 (2 H, m, ArCH_2C), 3.10, 3.37, 3.45, 3.82 (1 H, m, CH_2O), 4.30 (1 H, br m, CHN), 4.40 (1 H, br s, NH), 4.53 (1 H, m, OCHO), 5.04 (2 H, s, ArCH_2O), 5.45 (2 H, m, $\text{CH}=\text{CH}$), 6.90 (2 H, d, $J = 9$ Hz, para Ar), 7.11 (2 H, d, $J = 9$ Hz, para Ar), 7.3–7.5 (5 H, m, Ar); IR (CHCl_3) 3440, 3040, 3010, 2940, 2865, 1700, 1608, 1578, 1505, 1492, 1451, 1367, 1165, 1118, 1076, 1061, 1038, 972, 907, 868 cm^{-1} ; LRMS, m/e 446, 395, 379, 338, 326, 324, 322, 320, 310, 308, 292, 282, 280, 279, 270, 268, 256, 254, 242, 214, 197, 196, 176, 158, 142, 140, 127, 117, 115, 107, 101, 98, 97, 96, 92, 91 (100), 86, 85, 67, 65, 57, 56; $[\alpha]_D^{20} +6.6^\circ$ (c 0.35 g/mL, CHCl_3).

5(S)-[(tert-Butoxycarbonyl)amino]-2(R)-methyl-6-[4-(phenylmethoxy)phenyl]-trans-3-hexenoic acid, Protected

TyrAla Isostere 18. Method 1, Direct Oxidation. A solution of 0.400 g (0.807 mmol) of THP ether 17 in 50 mL of acetone was cooled to 0°C and treated dropwise with 1.40 mL (2.7 mmol) of Jones reagent (1.92 M). After 3 h, the mixture was diluted with water and extracted with several portions of ether. The ethereal fractions were extracted with five portions of 5% aqueous NaOH and the resulting aqueous extracts were combined and acidified to pH 2 with 10% aqueous HCl. Extraction of the acidified aqueous extracts with ether, drying (MgSO_4), concentration in vacuo (50°C), and recrystallization (1:2 ethyl acetate-hexanes) afforded the acid 18, 219 mg (66%), as a white solid; $^1\text{H NMR}$ (500 MHz) δ 1.25 (3 H, d, $J = 8$ Hz, CH_3) [other diastereoisomer; δ 1.22 (3 H, d, $J = 8$ Hz, CH_3)], 1.41 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.80 (2 H, m, ArCH_2C), 3.12 (1 H, m, CHCOO), 4.30 (1 H, m, CHN), 4.50 (1 H, br s, NH), 5.03 (2 H, s, ArCH_2O), 5.54 (1 H, dd, $J = 5$ and 16 Hz, $\text{CH}=\text{C}$), 5.62 (1 H, dd, $J = 8$ and 16 Hz, $\text{CH}=\text{C}$) [other isomer, δ 5.53 and 5.58 (same J values)], 6.90 (2 H, d, $J = 9$ Hz, para Ar), 7.08 (2 H, d, $J = 9$ Hz, para Ar), 7.3–7.45 (5 H, m, Ar); $^{13}\text{C NMR}$ (50 MHz) δ 16.9 (CH_3CH), 28.1 (CH_3CO), 40.7 (CHN), 42.1 (CH_2Ar), 52.8 (CHCOOH), 69.7 (ArCH_2O), 79.6 (CCH_3), 114.5 (Ar CCH_2CN), 127.3, 127.7, 128.3, 129.1, 129.7, 130.4, 131.6 (Ar and $\text{C}=\text{C}$), 137.0 ($\text{CHNC}=\text{C}$), 156.2 ($\text{O}=\text{CN}$), 157.3 (Ar CO), 179.4 (COOH); IR (CHCl_3) 3500–2400, 3435, 3040, 2975, 2930, 2870, 1703, 1609, 1582, 1492, 1452, 1391, 1268, 1240, 1167, 1075, 1061, 1020, 971, 863 cm^{-1} ; LRMS, m/e 425 (M^+), 352, 309, 308, 272, 248, 228, 197, 173, 172, 145, 129, 128, 117, 115, 107, 104, 91 (100), 85, 83 (96), 77, 65, 57; $[\alpha]_D^{20} +1.0^\circ$ (c 0.023 g/mL, CHCl_3); Anal. C, H, N.

Oxazolidone 19. A solution of 6.0 g (53 mmol) of 4-methyl-3-pentenoic acid in 20 mL of thionyl chloride and 30 mL of benzene was refluxed for 2 h. The volatiles were removed in vacuo (40°C) to give the crude acid chloride as a yellow oil: $^1\text{H NMR}$ (500 MHz) δ 1.11 (3 H, s, CH_3), 1.12 (3 H, s, CH_3), 3.55 (2 H, d, $J = 7$ Hz, CH_2), 5.27 (1 H, m, $\text{CH}=\text{C}$). In a separate flask, 6.0 g (33.9 mmol) of 5(S)-phenyl-4(R)-methyl-1,3-oxazolid-2-one in 80 mL of THF was cooled to -78°C , treated with 14.1 mL (31 mmol) of $n\text{-BuLi}$ (2.2 M in hexanes), and stirred at -78°C for 0.5 h. The acid chloride was added, and the mixture was stirred at -78°C for 0.5 h. The yellow solution was allowed to warm to 25°C and was quenched with saturated aqueous NaCl. Extraction with ether, drying (MgSO_4), and concentration in vacuo, followed by chromatography on silica gel (25% ethyl acetate-hexanes) afforded oxazolidone 19 as a white solid (6.49 g, 77%). The material obtained was (like the starting carboxylic acid) a mixture of α,β - and β,γ -unsaturated carbonyl compounds (27:73, determined by NMR). Oxazolidone 19: $^1\text{H NMR}$ (500 MHz) δ 0.90 (3 H, d, $J = 7$ Hz, CH_3CHN), 1.69 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 1.77 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 3.68 (2 H, d, $J = 7$ Hz, CH_2), 4.76 (1 H, m, CHCH_3), 5.38 (1 H, m, $\text{CH}=\text{C}$), 5.66 (1 H, d, $J = 7$ Hz, ArCH), 7.2–7.5 (5 H, m, Ar); IR (NaCl) 3040, 2970, 2938, 2920, 2880, 1781, 1700, 1632, 1455, 1371, 1352, 1235, 1192, 1178, 1122, 1068, 1042, 991 cm^{-1} ; LRMS: m/e 173 (M^+), 230, 178, 177, 160, 134, 119, 118, 117, 115, 105, 97, 96 (100), 95, 91, 86, 84, 81, 77, 70, 69, 67, 53.

Oxazolidone 20. A solution of 0.66 mL (4.7 mmol) of diisopropylamine in 3.5 mL of THF was cooled to 0°C and treated with 2.09 mL (4.6 mmol) of $n\text{-BuLi}$ (2.2 M in hexane). After being stirred for 0.5 h at 0°C , the solution was cooled to -78°C , and 1.0 g (3.65 mmol) of the oxazolidone 19 in 1 mL of THF was added dropwise. The mixture was kept at -78°C for 0.5 h followed by dropwise addition of 0.86 mL (7.2 mmol) of benzyl bromide. After 1 h at -78°C , the solution was warmed to -20°C over 1.5 h and then to 0°C over 1 h. Addition of saturated aqueous NH_4Cl , acidification with 5% aqueous HCl, extraction with ether, drying (MgSO_4) of the organic extract, and concentration in vacuo followed by chromatography on silica gel (5% ethyl acetate-hexanes) afforded the oxazolidone 20 as a colorless liquid (625 mg, 65% based on purity of oxazolidone 19, 88% diastereoisomeric excess by $^1\text{H NMR}$ analysis), which solidified upon standing at 25°C . A more polar fraction was eluted and identified as the unreacted α,β -unsaturated isomer of 19 (99 mg). 20: $^1\text{H NMR}$ (500 MHz) δ 0.68 (3 H, d, $J = 7$ Hz, CH_3CHN), 1.48 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 1.70 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 2.75 (1 H, dd, $J = 8$ and 13 Hz, ArCH_2), 3.09 (1 H, dd, $J = 8$ and 13 Hz, ArCH_2), 4.69 (1 H, m, CH_3CHN), 5.09 (1 H, m, CHCOH), 5.25 (1 H, d, $J = 10$ Hz, $\text{CH}=\text{C}$), 5.58 (1 H, d, $J = 7$ Hz, ArCHO), 7.2–7.5 (10 H, m, Ar); IR (CHCl_3) 3070, 3040, 2970, 2930, 2860, 1776, 1688, 1600, 1491, 1457, 1346,

1182, 1120, 1031, 942, 887, 872 cm^{-1} ; LRMS, m/e 273, 233, 232, 222, 220, 204, 178, 174, 148, 147, 145, 132, 131, 130, 129, 128, 117, 115, 104, 101, 92, 91, 85 (100), 79, 77, 67, 65, 57, 55; $[\alpha]_{\text{D}}^{20}$ -2.7° (as a 94:6 mixture of diastereoisomers); Anal. C, H, N.

4-Methyl-2(R)-(phenylmethyl)-3-penten-1-ol, Homoallylic Alcohol 21. A solution of 0.500 g (1.37 mmol) of oxazolidone 20 in 7 mL of THF was cooled to -78°C and treated dropwise with 1.37 mL (1.37 mmol) of LiAlH_4 (1 M in THF). The mixture was stirred at -78°C for 0.5 h and allowed to warm to 0°C over 1 h. After 0.5 h at 0°C , the clear solution was cooled to -78°C , and 1 mL of ethyl acetate was added. The reaction mixture was quenched with saturated aqueous NH_4Cl at -78°C and, after warming to 25°C , diluted with water. Extraction with ether, drying (MgSO_4), and concentration in vacuo (40°C), followed by chromatography on silica gel (25% ethyl acetate-hexanes), yielded the alcohol 21 as a colorless oil (0.235 g, 90%). 4(S)-Phenyl-5-(R)-methyloxazolidone was recovered (eluent, ethyl acetate) in 70% yield. 21: ^1H NMR (500 MHz) δ 1.42 (3 H, s, CH_3), 1.5 (1 H, br s, OH), 1.70 (3 H, s, CH_3), 2.51 (1 H, dd, $J = 7$ and 14 Hz, ArCH_2), 2.69 (1 H, dd, $J = 7$ and 14 Hz, ArCH_2), 2.77 (1 H, m, CH), 3.40 (1 H, dd, $J = 8$ and 10 Hz, CH_2O), 3.57 (1 H, dd, $J = 5$ and 10 Hz, CH_2O), 4.94 (1 H, d, $J = 10$ Hz, $=\text{CH}$), 7.1–7.3 (5 H, m, Ar); IR (NaCl) 3380, 3090, 3060, 3030, 2970, 2930, 2860, 1860, 1500, 1450, 1070, 1030 cm^{-1} ; LRMS, m/e 190 (M^+), 130, 117, 99, 98, 92, 91, 86, 85, 84, 83, 82, 81, 79, 77, 71, 69, 57 (100), 55; $[\alpha]_{\text{D}}^{20}$ -20.9° (c 0.12 g/mL, CHCl_3).

2-Methyl-4(R)-(phenylmethyl)-5-[(2-tetrahydropyran-2-yl)oxy]-2-pentene, THP Ether 22. Alcohol 21 (0.200 g) was converted to the THP ether 22 as a colorless liquid (after chromatography with 25% ethyl acetate-hexanes) (0.299 g, 100%) as described above for the synthesis of methyl 3-[(2-tetrahydropyran-2-yl)oxy]-2(S)-methylpropionate. 22: ^1H NMR (500 MHz) δ 1.41 (3 H, d, $J = 5$ Hz, CH_3), 1.5–1.9 (6 H, m, THP ether), 1.64 (3 H, s, CH_3), 2.50 (1 H, m, ArCH_2), 1.75–2.97 (2 H, ArCH_2 and $\text{CHC}=\text{C}$), 3.25 (1 H, m, CH_2O), 3.64 (1 H, m, CH_2O), 3.49 (1 H, m, CH_2O), 3.87 (1 H, m, CH_2O), 4.56 (1 H, m, OCHO), 4.99 (1 H, m, $\text{CH}=\text{C}$), 7.1–7.3 (5 H, m, Ar); IR (NaCl) 3065, 2930, 2860, 1738 (w), 1492, 1451, 1350, 1201, 1138, 1119, 1078, 1031, 975, 908, 870, 815, 742, 699 cm^{-1} ; LRMS, m/e 274 (M^+), 244, 214, 175, 174, 158, 157, 143, 131, 129, 128, 118, 117, 115, 101, 91, 85 (100), 84, 77, 67, 57, 55; $[\alpha]_{\text{D}}^{20}$ -27.40° (c 0.16 g/mL in CHCl_3); Anal. C, H.

2(S)-(Phenylmethyl)-3-[(2-tetrahydropyran-2-yl)oxy]-propionaldehyde, Aldehyde 23. A solution of 1.20 g (4.37 mmol) of 22 in 15 mL of $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$ (5:1) was cooled to -78°C . A stream of ozone was passed through the solution until a faint blue color was observed (5 min). The mixture was quenched with excess dimethyl sulfide at -78°C , followed by warming to 25°C and concentration in vacuo (25°C). The resulting colorless oil was normally used in the next step without further purification (the only major impurity being dimethyl sulfoxide). Chromatography on silica gel (25% ethyl acetate-hexanes) afforded pure aldehyde 23 as a clear liquid (0.78 g, 74%): ^1H NMR (500 MHz) δ 1.4–1.8 (6 H, m, THP ether), 2.83 (2 H, m, CHCHO and ArCH_2), 3.07 (1 H, m, ArCH_2), 3.45–3.6 (2 H, m, OCH_2), 3.8 (1 H, m, OCH_2), 3.95–4.05 (1 H, m, OCH_2), 4.55 (1 H, dt, OCHO), 7.2–7.4 (5 H, m, Ar), 9.93 (1 H, $2 \times$ d, $J = 1$ Hz, CHO); IR (NaCl) 3090, 3060, 3040, 2940, 2880, 2830, 2780, 1725, 1500, 1470, 1455, 1360, 1350, 1328, 1135, 1080, 1040, 1000, 973, 908, 872, 812 cm^{-1} ; LRMS, m/e 248 (M^+), 157, 145, 139, 133, 131, 129, 128, 91, 86, 85 (100), 84, 79, 78, 77, 76, 67, 65, 57, 55; $[\alpha]_{\text{D}}^{20}$ $+42.8^\circ$ (c 0.111 g/mL, CHCl_3).

2-(2-Methylpropyl)-4-methyl-3-pentenoic Acid, 24a. At 0°C 45.5 mL (100 mmol) of *n*-BuLi (2.2 M in hexane) was added to a solution of 14.5 mL (105 mmol) of diisopropylamine in 75 mL of THF. After being stirred at 0°C for 0.5 h, the pale yellow solution was cooled to -78°C , treated with 5.0 g (43 mmol) of 4-methyl-3-pentenoic acid, stirred at -78°C for 0.5 h, warmed to 25°C for 0.5 h, and finally cooled to -78°C , where 11.5 mL (100 mmol) of isobutyl iodide was added. The mixture was stirred at -78°C for 0.5 h, warmed to 25°C over 2 h, and stirred 1 h. Quenching with 5% HCl and extracted with ether, followed by drying (MgSO_4) and concentration in vacuo, afforded the crude product, which was chromatographed on silica gel (25% ethyl acetate-hexanes) to yield 4.0 g (56%) of the acid 24a as a colorless oil, which solidified upon standing at 25°C : ^1H NMR (500 MHz) δ 0.89 (3 H, d, $J = 6$ Hz, CH_3 (*i*-Bu)), 0.93 (3 H, d, $J = 6$ Hz, CH_3

(*i*-Bu)), 1.37 (1 H, m, CH (*i*-Bu)), 1.61 (2 H, m, CH_2), 1.69 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 1.74 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 3.32 (1 H, m, $\text{CHC}=\text{C}$), 5.08 (1 H, d, $J = 7$ Hz, $\text{CH}=\text{C}$); IR (NaCl) 3500–2500, 2980, 2860, 1704, 1650, 1469, 1440, 1412, 1388, 1378, 1370, 1290, 1212, 1190, 1111, 939, 842, 829, 805, 680 cm^{-1} ; LRMS, m/e 170 (M^+), 153, 137, 127, 125, 115, 114, 99, 98, 97, 96, 95, 86, 85, 84, 83, 82, 81, 70, 69 (100), 67, 59, 56, 55, 53, 51.

2-(2-Methylpropyl)-4-methyl-3-penten-1-ol, Alcohol 25a. A solution of 3.0 g (17.6 mmol) of acid 24a in 10 mL of THF was added dropwise at 0°C to a slurry of 1.0 g (26.4 mmol) of LiAlH_4 in 30 mL of THF. The mixture was stirred at 25°C for 1 h and warmed to 50°C for 2 h. Excess 5% aqueous HCl was carefully added at 0°C and the resulting solution was extracted several times with ether. Drying (MgSO_4) and concentration in vacuo (50°C) afforded 2.70 g (98% of the alcohol 25a as a colorless liquid: ^1H NMR (500 MHz) δ 0.85 (3 H, d, $J = 7$ Hz, CH_3CH), 0.89 (3 H, d, $J = 7$ Hz, CH_3CH), 1.11 (2 H, m, CH_2 -*i*-Pr), 1.5 (1 H, br s, OH), 1.55 (1 H, m, CHCH_3), 1.68 (3 H, d, $J = 1$ Hz, $\text{CH}_3\text{C}=\text{C}$), 1.75 (3 H, d, $J = 1$ Hz, $\text{CH}_3\text{C}=\text{C}$), 2.58 (1 H, m, $\text{CHC}=\text{C}$), 3.26 (1 H, dd, $J = 8$ and 11 Hz, CH_2OH), 3.50 (1 H, dd, $J = 5$ and 11 Hz, CH_2OH), 4.81 (1 H, d, $J = 9$ Hz, $\text{CH}=\text{C}$); IR (NaCl) 3350, 2980, 2870, 1670, 1470, 1450, 1385, 1370, 1170, 1055, 1020, 840 cm^{-1} ; LRMS, m/e 156 (M^+), 125, 109, 81, 69 (100), 67, 57, 55.

2,6-Dimethyl-4-[(2-tetrahydropyran-2-yl)oxy]methyl]-2-heptene, THP Ether 26a. Alcohol 25a (3.2 g) was converted to the THP ether 26a (4.88 g, 99%) as a colorless liquid (after chromatography with 25% ethyl acetate-hexanes) as described above for the synthesis of methyl 3-[(2-tetrahydropyran-2-yl)oxy]-2(S)-methylpropionate. 26a: ^1H NMR (500 MHz) δ 0.86 (3 H, d, $J = 7$ Hz, CH_3CH), 0.91 (3 H, d, $J = 7$ Hz, CH_3CH), 1.28 (2 H, m, CH_2 -*i*-Pr), 1.45–1.85 (7 H, m, THP ether and $\text{CH}(\text{CH}_3)_2$), 1.68 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 1.73 (3 H, s, $\text{CH}_3\text{C}=\text{C}$), 2.61 (1 H, m, $\text{CHC}=\text{C}$), 3.20 (1 H, m, CH_2O), 3.50 (2 H, m, CH_2O), 3.83 (1 H, m, CH_2O), 4.54 (1 H, m, OCHO), 4.76 (1 H, m, $\text{CH}=\text{C}$); IR (NaCl) 2980, 2870, 1468, 1452, 1441, 1385, 1377, 1368, 1352, 1342, 1322, 1282, 1261, 1200, 1183, 1139, 1120, 1078, 1030, 980, 909, 871, 819 cm^{-1} ; LRMS, m/e 240 (M^+), 169, 157, 140, 139, 126, 124, 123, 115, 109, 102, 101, 95, 86, 85 (100), 83, 82, 81, 69, 67, 57, 55; HRMS, m/e 240.2104 (calcd 240.2119).

4-Methyl-2-[(2-tetrahydropyran-2-yl)oxy]methyl]pentanal, Aldehyde 27a. THP ether 26a (2.0 g, 8.3 mmol) was ozonized as described above for the preparation of aldehyde 23, to yield 1.55 g (87%) of aldehyde 27a after chromatography (25% ethyl acetate-hexanes) on silica gel: ^1H NMR (500 MHz) δ 0.92 (6 H, m, CH_3), 1.30 (2 H, m, CH_2), 1.42–1.82 (7 H, m, THP ether and $\text{CH}(\text{CH}_3)_2$), 2.63 (1 H, m, CHCHO), 3.45–4.02 (4 H, m, $2 \times \text{CH}_2\text{O}$), 4.58 (1 H, m, OCHO), 9.690 and 9.692 (1 H, $2 \times$ d, ratio 1:1, $J = 5$ Hz, CHO); IR (NaCl) 2970, 2870, 2720, 1726, 1468, 1454, 1442, 1388, 1371, 1354, 1204, 1125, 1080, 1038, 908, 872, 819 cm^{-1} ; LRMS, m/e 214 (M^+), 157, 139, 129, 114, 113, 102, 101, 95, 86, 85 (100), 83, 71, 69, 67, 57, 56, 55, 53.

5(S)-[(*tert*-Butoxycarbonyl)amino]-6-phenyl-2(R)-(phenylmethyl)-1-[(2-tetrahydropyran-2-yl)oxy]-*trans*-3-hexene, THP ether 28a: ^1H NMR (500 MHz) δ 1.40 (9 H, s, $t\text{-C}_4\text{H}_9$), 1.35–1.90 (6 H, m, THP ether), 2.4–2.95 (5 H, m, ArCH_2 and CHCH_2Ar), 3.25, 3.50, 3.68, and 3.82 (each 1 H, m, CH_2O), 4.33 (2 H, br m, NH and CHN), 4.54 (1 H, m, OCHO), 5.26 (1 H, m, $\text{CH}=\text{C}$), 5.45 (1 H, m, $\text{CH}=\text{C}$), 7.05–7.35 (10 H, m, Ar); LRMS, m/e 374, 318, 305, 304, 273, 272, 256, 249, 248, 234, 230, 227, 218, 216, 204, 200, 186, 178, 177, 174, 173, 172, 162, 161, 160, 144, 143, 142, 136, 134, 130, 129, 122, 118, 117, 115, 107, 105, 101, 97, 96, 92, 91, 89, 86, 85 (100), 84, 77, 67, 57, 51; $[\alpha]_{\text{D}}^{20}$ -24.5° (c 0.05 g/mL, CHCl_3); Anal. C, H, N; HRMS (CI), m/e 465.2930 (calcd 465.2879).

5(S)-[(*tert*-Butoxycarbonyl)amino]-7-methyl-2(R)-(phenylmethyl)-1-[(2-tetrahydropyran-2-yl)oxy]-*trans*-3-octene, THP ether 28b: ^1H NMR (500 MHz) δ 0.88 (6 H, m, CH_3CH), 1.15–1.9 (9 H, m, THP ether and $\text{CH}_2\text{CH}(\text{CH}_3)_2$), 1.44 (9 H, s, $t\text{-C}_4\text{H}_9$), 2.55 (2 H, m, ArCH_2) and CHCH_2Ar), 2.85 (1 H, m, ArCH_2), 5.30, 5.51, 5.67, and 5.82 (each 1 H, m, CH_2O), 4.05 (1 H, m, CHN), 4.26 (1 H, br s, NH), 4.55 (1 H, m, OCHO), 5.20 (1 H, m, $\text{CH}=\text{C}$), 5.49 (1 H, m, $\text{CH}=\text{C}$), 7.2–7.3 (5 H, m, Ar); IR (CHCl_3) 3340, 3085, 3058, 3025, 2960, 2870, 1705, 1601, 1510, 1496, 1455, 1390, 1368, 1248, 1201, 1170, 1122, 1078, 1032, 975, 908, 871, 825, 755, 701 cm^{-1} ; LRMS, m/e 291, 290, 274, 261, 246, 234, 230, 218, 201, 174, 173, 172, 164, 159, 157, 155, 144, 138, 130, 129, 117, 115,

112, 101, 100, 96, 92, 91, 85, 67, 57 (100); $[\alpha]_D^{20}$ -33.7° (c 0.1 g/mL, CHCl₃).

4(S)-[(*tert*-Butoxycarbonyl)amino]-2,9-dimethyl-7-[[2-tetrahydropyranyl]oxy]methyl]-*trans*-5-decene, THP Ether 28c. Diastereoisomer I: ¹H NMR (500 MHz) δ 0.83-0.91 (6 H, m, CH₃), 1.15-1.85 (9 H, m, THP ether and CH₂CH(CH₃)₂), 1.42 (9 H, s, *t*-C₄H₉), 2.36 (1 H, m, CHC=), 3.22, 3.48, 3.58, and 3.83 (each 1 H, m, CH₂O), 4.08 (1 H, m, CHN), 5.30 (1 H, br s, NH), 4.57 (1 H, m, OCHO), 5.25-5.45 (2 H, m, CH=CH). Diastereoisomer II: ¹H NMR (500 MHz) δ 0.82-0.91 (6 H, m, CH₃), 1.1-1.9 (9 H, m, THP ether and CH₂CH(CH₃)₂), 1.42 (9 H, s, *t*-C₄H₉), 2.38 (1 H, m, CHC=), 3.26, 3.45, 3.57, and 3.85 (each 1 H, m, CH₂O), 4.09 (1 H, m, CHN), 4.32 (1 H, br s, NH), 4.55 (1 H, m, OCHO), 5.35-5.43 (2 H, m, CH=CH). Diastereoisomers I and II were virtually indistinguishable by IR and LRMS: IR (NaCl) 3340, 2960, 2875, 1700, 1510, 1468, 1452, 1386, 1368 cm⁻¹; LRMS, *m/e* 340, 312, 284, 283, 257, 256, 240, 227, 200, 196, 182, 166, 140, 138, 130, 123, 110, 95, 86, 85 (100); diastereoisomer I, $[\alpha]_D^{20}$ -27.6° (c 0.1 g/mL, CHCl₃); diastereoisomer II, $[\alpha]_D^{20}$ +0.7° (c 0.1 g/mL, CHCl₃).

5(S)-[(*tert*-Butoxycarbonyl)amino]-6-phenyl-2(R)-(phenylmethyl)-*trans*-3-hexenoic Acid, protected PhePhe isostere 29a: ¹H NMR (500 MHz) δ 1.40 (9 H, s, *t*-C₄H₉), 2.76 (3 H, m, ArCH₂CHN and ArCH₂CHCO), 3.08 (1 H, m, ArCH₂CHCO), 3.29 (1 H, m, CHCOO), 4.35-4.55 (2 H, br m, CHNH), 5.44 (1 H, dd, *J* = 7 and 16 Hz, CH=C), 5.58 (1 H, dd, *J* = 10 and 16 Hz, CH=C), 7.05-7.35 (10 H, m, Ar); ¹³C NMR (50 MHz) δ 28.3 (CH₃CO), 38.3, (CHN), 41.6 (ArCH₂CN), 50.5 (ArCH₂CHCOO), 52.7 (CHCOOH), 79.6 (OCCH₃), 126.4, 127.1, 127.4, 128.3, 129.1, 129.5, 130.1 (Ar and C=C), 133.5 (Ar CCH₂CN), 137.2 (CHC=C), 138.4 (Ar CCH₂CCOOH), 155.1 (O=CN), 178.4 (COOH); IR (CHCl₃) 3500-2500, 3440, 3040, 2980, 2940, 2860, 1705, 1685, 1600, 1492, 1453, 1369, 1286, 1165, 1075, 1031, 972, 922 cm⁻¹; LRMS, *m/e* 318, 317, 305, 304, 249, 248, 234, 230, 214, 204, 200, 187, 186, 170, 161, 143, 130, 129, 128, 117, 115, 105, 97, 95, 92, 91 (100), 85, 81, 77, 65, 57, 55; $[\alpha]_D^{20}$ -17.0° (c 0.1 g/mL, CHCl₃); HRMS (CI), *m/e* 395.2107 (calcd 395.2104).

5(S)-[(*tert*-Butoxycarbonyl)amino]-7-methyl-2(R)-(phenylmethyl)-*trans*-3-octenoic Acid, Protected LeuPhe Isostere 29b. Method 2, Hydrolysis/Oxidation. A solution of 0.1313 g (0.3 mmol) of the THP ether 29a and 0.5 mg of pyridinium *p*-toluenesulfonate in 5 mL of CH₃OH was stirred at 25 °C for 10 h. The volatiles were removed in vacuo, and the residue was dissolved in 5 mL of acetone and cooled to 0 °C. Jones reagent (2.5 mL, 4.8 mmol, 1.92 M) was added, and the mixture was stirred 1 h at 0 °C. Ether and water were added, and the layers were separated. The aqueous layer was extracted with several portions of fresh ether, and the product was recovered by extraction of the combined ethereal layers with 5% aqueous NaOH. The aqueous extracts were acidified with 10% aqueous HCl and extracted with ether. Drying (MgSO₄) followed by concentration in vacuo (50 °C) afforded 68.5 mg (63%) of 29b

as a pale yellow oil: ¹H NMR (500 MHz) δ 0.86 (3 H, d, *J* = 6 Hz, CH₃), 0.87 (3 H, d, *J* = 6 Hz, CH₃), 1.23 (2 H, m, CH₂-*i*-Pr), 1.44 (9 H, s, *t*-C₄H₉), 1.54 (1 H, m, CH(CH₃)₂), 2.81 (1 H, dd, *J* = 7 and 13 Hz, ArCH₂), 3.11 (1 H, dd, *J* = 7 and 13 Hz, ArCH₂), 3.28 (1 H, m, CHCOO), 4.05 (1 H, m, CHN), 4.28 (1 H, s, NH), 5.35 (1 H, dd, *J* = 6 and 15 Hz, CH=), 5.63 (1 H, dd, *J* = 8 and 15 Hz, CH=), 7.14-7.28 (5 H, m, Ar); ¹³C NMR (50 MHz) δ 2.5 (CH₃CH), 24.5 (CHCH₃), 28.3 (CH₃CO), 38.4 (CHN), 44.4 (C-H₂CHN), 50.4 (ArCH₂), 65.8 (CHCOOH), 79.8 (OCCH₃), 126.4, 126.5 (Ar para C and CHC=C), 128.3 (Ar ortho C), 129.1 (Ar meta C), 134.9 (CHC=C), 138.4 (Ar CCH₂), 155.3 (O=CN), 178.2 (COOH); IR (CHCl₃) 3500-2500, 3440, 3080, 3040, 2960, 2935, 2875, 1703, 1650, 1502, 1492, 1452, 1392, 1370, 1282, 1165, 1067, 972, 912, 870 cm⁻¹; LRMS, *m/e* 304, 260, 259, 249, 248, 244, 205, 204, 200, 161, 156, 144, 143, 130, 117, 115, 112, 105, 96, 92, 91 (100), 86, 84, 77, 65, 59, 57; $[\alpha]_D^{20}$ -32.1° (c 0.025 g/mL, CHCl₃); HRMS (CI), *m/e* 361.2222 (calcd 361.2261).

5(S)-[(*tert*-Butoxycarbonyl)amino]-7-methyl-2-(methylpropyl)-*trans*-3-octenoic Acid, Protected LeuLeu Isostere 29c. Diastereoisomer I: ¹H NMR (500 MHz) δ 0.87 (1.5 H, d, *J* = 6 Hz, CH₃), 0.90 (4.5 H, m, CH₃), 1.34 (1 H, m, CH(CH₃)₂), 1.43 (9 H, s, *t*-C₄H₉), 1.63 (2 H, m, CH₂-*i*-Pr), 3.09 (1 H, m, CHCOO), 4.12 (1 H, br m, CHN), 4.40 (1 H, br s, NH), 5.54 (2 H, m, CH=CH). Diastereoisomer II: ¹H NMR (500 MHz) δ 0.87 (1.5 H, d, *J* = 6 Hz, CH₃), 0.90 (4.5 H, m, CH₃), 1.34 (1 H, m, CH(CH₃)₂), 1.44 (9 H, s, *t*-C₄H₉), 1.62 (2 H, m, CH₂-*i*-Pr), 3.07 (1 H, m, CHCOO), 4.10 (1 H, br m, CHN), 4.40 (1 H, br s, NH), 5.46 (1 H, dd, *J* = 6 and 16 Hz, CH=), 5.56 (1 H, dd, *J* = 8 and 16 Hz, CH=). The ¹³C NMR, MS, and IR spectra for both diastereoisomers were virtually indistinguishable: ¹³C NMR (50 MHz) δ 22.4, 22.6 (2 × CH₃CH), 24.7, 25.5 (2 × CHCH₃), 28.4 (CH₃CO), 41.3 (CHN), 44.6 (CH₂CN), 47.0 (CH₂CCOOH), 50.2 (CHCOOH), 79.6 (OCCH₃), 127.7 (C=C), 134.3 (C=C), 155.3 (O=CN), 179.7 (COOH); IR (CHCl₃) 3500-2500, 3440, 2960, 2875, 1702, 1492, 1468, 1453, 1389, 1320, 1280, 1240; 1168, 1076, 1062, 1051, 970, 870 cm⁻¹; LRMS, *m/e* 272, 271, 270, 226, 225, 215, 184, 182, 170, 157, 156, 154, 152, 130, 117, 112, 109, 105, 96, 95, 91, 86, 85, 84, 82, 77, 69, 67, 57 (100); HRMS (CI), *m/e* 327.2409 (calcd 327.2418); diastereoisomer I, $[\alpha]_D^{20}$ +18.1° (c 0.05 g/mL, CHCl₃); diastereoisomer II, $[\alpha]_D^{20}$ -29.3° (c 0.02 g/mL, CHCl₃).

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New Spongiane Diterpenes from an Australian Nudibranch

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Nine new spongiane-type diterpenes have been isolated from a nudibranch collected in South Australia and tentatively identified as *Ceratosoma brevicaudatum* (Abraham). Variation in extent and site of oxidation distinguishes these diterpenes from others of this skeletal class. Structures were determined by detailed spectroscopic analyses with emphasis on ¹H and ¹³C NMR data. Relayed coherence transfer and long-range COSY ¹H NMR experiments were used to identify spin systems of partial structures involving unresolved, overlapping signals.

A family of diterpenes sharing a common skeleton represented by 1¹ and designated spongianes² have been re-

ported from various sponge sources.^{3,4} Metabolites with rearranged versions of this skeleton have been isolated